

# Product Sheet

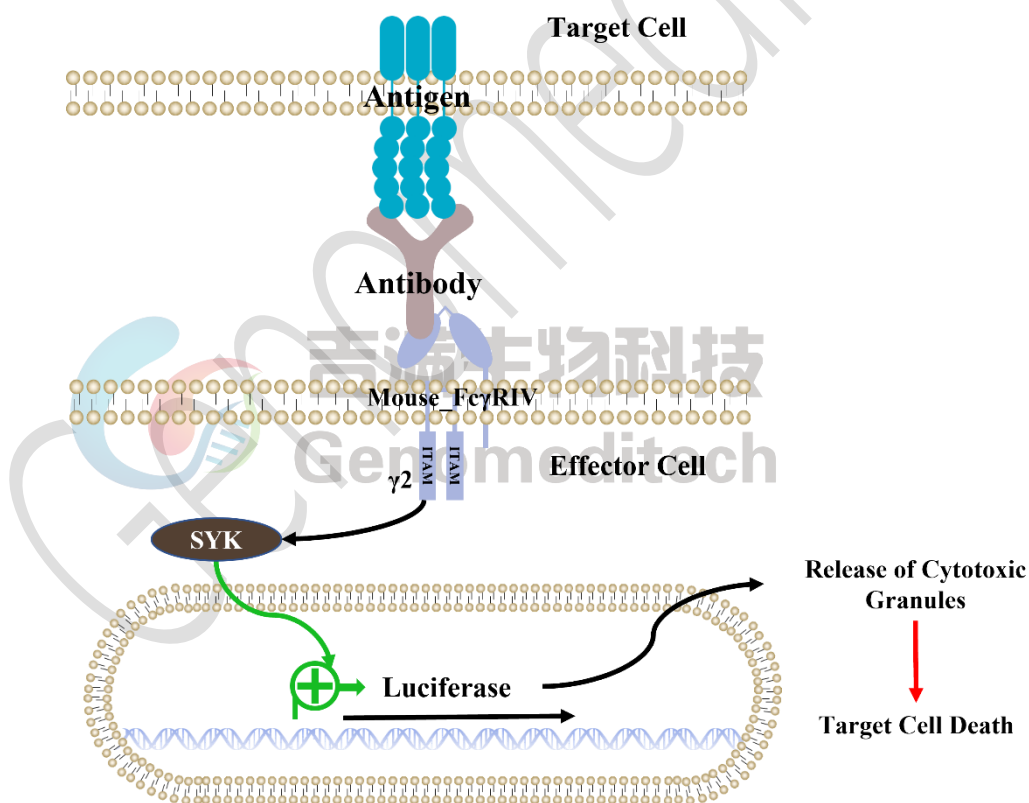
## ADCC M\_FcγRIV Jurkat Effector Cell Line

Catalog number: GM-C09105

Version 3.3.1.241226

ADCC, or antibody-dependent cell-mediated cytotoxicity, refers to the process by which immune cells expressing Fc receptors directly kill target cells that specifically bind to antibodies through recognition of the Fc region of the antibodies. Nowadays, the mechanism of ADCC is used to detect and evaluate the efficacy of antibodies or target cells. Antibodies bind to target antigens on the cell surface, if the Fc region of the antibody simultaneously binds to the FcγRIV receptor on the surface of effector cells (mouse FcγRIV is the main receptor involved in mouse ADCC and is more similar to human FcγRIIIa), the two types of cells undergo multiple cross-linking, leading to the activation of the ADCC mechanism pathway.

ADCC M\_FcγRIV Jurkat Effector Cell Line is a clonal stable Jurkat cell line constructed using lentiviral technology, constitutive expression of the mouse FcγRIV gene, along with signal-dependent expression of a luciferase reporter gene. When IgG binds to target cells and effector cells, it leads to the expression of luciferase, which can be used to evaluate the biological activity of antibodies in the mechanism of ADCC.



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## Specifications

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<b>Quantity</b>	5E6 Cells per vial, 1 mL
<b>Product Format</b>	1 vial of frozen cells
<b>Shipping</b>	Shipped on dry ice
<b>Storage Conditions</b>	Liquid nitrogen immediately upon receipt

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<b>Recovery Medium</b>	RPMI 1640+10% FBS+1% P.S
<b>Growth medium</b>	RPMI 1640+10% FBS+1% P.S+3.5 µg/mL Blasticidin+0.75 µg/mL Puromycin
<b>Note</b>	None
<b>Freezing Medium</b>	90% FBS+10% DMSO
<b>Growth properties</b>	Suspension
<b>Growth Conditions</b>	37°C, 5% CO <sub>2</sub>

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<b>Mycoplasma Testing</b>	The cell line has been screened to confirm the absence of Mycoplasma species.
<b>Safety considerations</b>	Biosafety Level 2
<b>Note</b>	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

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## Materials

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<b>Reagent</b>	<b>Manufacturer/Catalogue No.</b>
RPMI 1640	VivaCell/C3010-0500
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/ <a href="#">GM-040404</a>
Puromycin	Genomeditech/ <a href="#">GM-040401</a>
H_ALPPL2(ALPG) HEK-293 Cell Line	Genomeditech/ <a href="#">GM-C26399</a>
Anti-ALPPL2 mIgG2a antibody(SGN-ALPV)	Genomeditech/ <a href="#">GM-58773AB</a>
GMOne-Step Luciferase Reporter Gene Assay Kit	Genomeditech/ <a href="#">GM-040503</a>

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## Figures

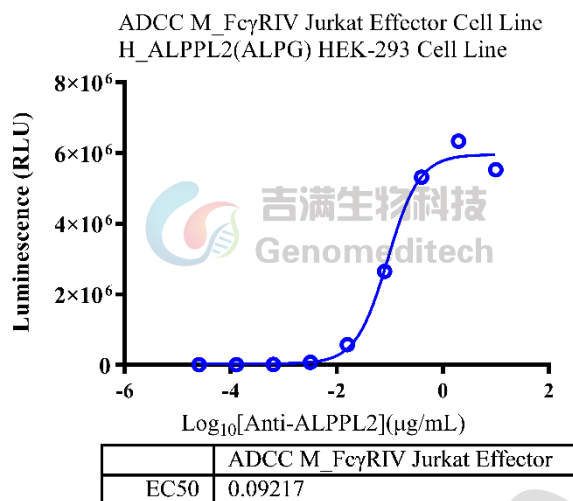


Figure 1 | Response to Anti-ALPPL2 mIgG2a antibody(SGN-ALPV). Serial dilutions of the Anti-ALPPL2 mIgG2a antibody(SGN-ALPV) (Cat. [GM-58773AB](#)) and 1.5E5 cells/well of the ADCC M\_FcγRIV Jurkat Effector Cell Line (Cat. GM-C09105) were added to 1.5E4 cells/well of H\_ALPPL2(ALPG) HEK-293 Cell Line (Cat. [GM-C26399](#)) for 6 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). The maximum induction fold was approximately[499.5]. Data are shown by drug mass concentration.

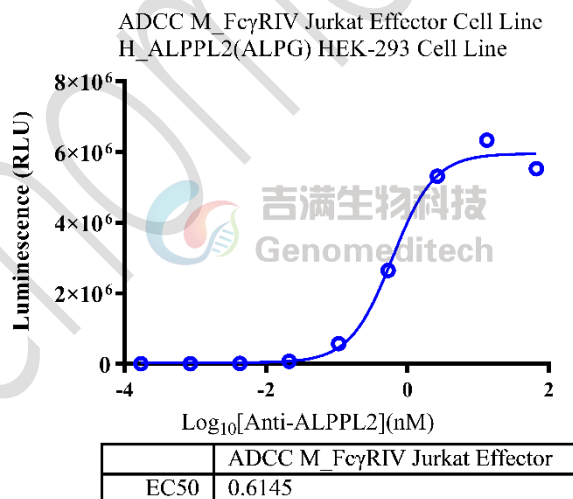


Figure 2 | Response to Anti-ALPPL2 mIgG2a antibody(SGN-ALPV). Serial dilutions of the Anti-ALPPL2 mIgG2a antibody(SGN-ALPV) (Cat. [GM-58773AB](#)) and 1.5E5 cells/well of the ADCC M\_FcγRIV Jurkat Effector Cell Line (Cat. GM-C09105) were added to 1.5E4 cells/well of H\_ALPPL2(ALPG) HEK-293 Cell Line (Cat. [GM-C26399](#)) for 6 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). The maximum induction fold was approximately[499.5]. Data are shown by drug molar concentration.

## Cell Recovery

Recovery Medium: RPMI 1640+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at  $-70^{\circ}\text{C}$ . Storage at  $-70^{\circ}\text{C}$  will result in loss of viability.

- a) Thaw the vial by gentle agitation in a  $37^{\circ}\text{C}$  water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium. And spin at approximately  $176 \times g$  for 5 minutes. Discard supernatant.
- d) Resuspend cell pellet with the recommended complete medium. And dispense the suspension into 1 - 2 T-25 culture flasks.
- e) Incubate the culture at  $37^{\circ}\text{C}$  in a suitable incubator. A 5%  $\text{CO}_2$  in air atmosphere is recommended if using the medium described on this product sheet.

## Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- a) Centrifuge at  $176 \times g$  for 3 minutes to collect cells.
- b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to  $5 \times 10^6$  cells/mL.
- c) Aliquot 1 mL into each vial.
- d) Place the vial in a controlled-rate freezing container and store at  $-80^{\circ}\text{C}$  for at least 1 day, then transfer to liquid nitrogen as soon as possible.

## Cell passage

Growth medium: RPMI 1640+10% FBS+1% P.S+3.5  $\mu\text{g}/\text{mL}$  Blasticidin+0.75  $\mu\text{g}/\text{mL}$  Puromycin

Approximately 48-72 hours after the initial thawing, the cells can be passaged for the first time. After this initial passage, the culture medium can be adjusted to growth medium supplemented with antibiotics. If cells are not passaged within 48 hours, it is recommended to add some fresh recovery medium and place the flask horizontally.

- a) When the cell density reaches  $1.5 - 2 \times 10^6$  cells/mL, subculture the cells. Do not allow the cell density to exceed  $2 \times 10^6$  cells/mL.
- b) It is recommended to use T-25 flasks for subculturing.
- c) These cells are suspension cells, and it is recommended to use the "half-medium change" method to maintain optimal cell conditions during passaging.
- d) During passaging, you can directly add fresh growth medium to the culture flask, gently pipette to resuspend the cells, and then transfer the cell suspension to a new T-25 flask for continued culture.

**Subcultivation Ratio: Maintain cultures at a cell concentraion between  $3 \times 10^5$  and  $1 \times 10^6$  viable cells/mL.**

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**Medium Renewal: Every 2 to 3 days**

## Notes

- These cells are sensitive to density, so please ensure that the cell density is maintained within an appropriate range during culture and subculturing.
- During the first passage, pay attention to the nutrient supply; if not subculturing, make sure to add fresh recovery medium every other day as needed.

## Related Products

FcγR	
<a href="#">Cynomolgus_FcRn MDCK Cell Line</a>	<a href="#">H_FCGR1A(CD64) CHO-K1 Cell Line</a>
<a href="#">H_FCGR1A(CD64) HEK-293 Cell Line</a>	<a href="#">H_FCGR2A(CD32A) CHO-K1 Cell Line</a>
<a href="#">H_FCGR2B(CD32B) CHO-K1 Cell Line</a>	<a href="#">H_FCGR3A(CD16a) 158F CHO-K1 Cell Line</a>
<a href="#">H_FCGR3A(CD16a) 158V CHO-K1 Cell Line</a>	<a href="#">H_FCGR3B(CD16b) CHO-K1 Cell Line</a>
<a href="#">H_FcRn CHO-K1 Cell Line</a>	<a href="#">H_FcRn MDCK Cell Line</a>
<a href="#">Mouse_FcRn MDCK Cell Line</a>	
<a href="#">Anti-FcRn hIgG4 Reference Antibody(Rozabio)</a>	<a href="#">Anti-H_FcRn IgG4 Antibody(Rozanolixizumab)</a>
<a href="#">Anti-Mouse CD1632 mIgG2b Antibody(2.4G2)</a>	
ADCCP	
<a href="#">ADCC FcγRIIIa(158F) Jurkat Effector Cell Line</a>	<a href="#">ADCC FcγRIIIa(158V) DDX35TM Jurkat Effector Cell Line</a>
<a href="#">ADCC FcγRIIIa(158V) Jurkat Effector Cell Line</a>	<a href="#">ADCP FcγRIIa DDX35TM Jurkat Effector Cell Line</a>
<a href="#">ADCP FcγRIIa Jurkat Effector Cell Line</a>	<a href="#">ADCP FcγRIIa R131 Jurkat Effector Cell Line</a>
<a href="#">ADCP FcγRIIb Jurkat Effector Cell Line</a>	

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